SUMMIT
PATHOLOGY

Specimen Submission Guide

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Table of Contents

Gynecologic Cytology and Affiliated Molecular Testing

Specimens submitted to Summit Pathology should be labeled with two separate and unique identifiers, as required by the College of American Pathologists (CAP). All specimens must be accompanied by a Summit Pathology requisition form. If you are out of forms, please go to our website, www.summitpathology.com and go to the "Forms" tab. Print the requisition labeled "Cytology/Molecular/AP Requisition". Requisition forms must be completed with patient name, date of birth, complete address and phone number, insurance information, date of specimen collection, physician name, appropriate clinical history, ICD9 code, site of specimen, and testing requested. Specimen containers should be placed into the main part of the plastic biohazard bags supplied by Summit Pathology. The requisition form should be placed into the outer pouch of the bag.

Pap Tests

Summit Pathology accepts the following Papanicolaou modalities: ThinPrep®, SurePath™ and conventional slide Pap tests. Each test involves a specific collection technique. Please reference the attachments at the end of this guide for proper collection technique. Please remember that use of lubricants can interfere with adequate sampling of the ThinPrep® Pap test. An additional attachment at the end of this guide explains this further. Pap specimens do not need to be refrigerated.

Attachment 1 - ThinPrep® Specimen Collection Guide
Attachment 2 – Lubricant Use with the ThinPrep® Pap test
Attachment 3 - SurePath™ Specimen Collection Guide

HPV

Summit Pathology uses the Roche COBAS HPV Test. This test for the Human Papillomavirus, which screens the following high risk sub-types: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66 and 68. In addition to the screening result, this test will provide positive or negative results individually for the 16 and 18 genotpyes. The specimen sample for this test can be taken from either the ThinPrep® or SurePath™ Pap vial. Summit Pathology keeps pap specimens in our lab for six weeks. HPV tests can be added at any time in the six week window.

Chlamydia & Gonorrhea

Summit Pathology uses the Gen-Probe® Aptima® II Combo Assay for Chlamydia and Gonorrhea testing. The specimen sample for this test can be taken from either the ThinPrep® or SurePath™ Pap vial. Those tests can be added to a pap that was sent to Summit Pathology for one month after the specimen was received.

However, if you are not taking a liquid based pap specimen, **or** the patient is male, there are two other options. A urine specimen may be submitted using the Gen-Probe® Aptima Urine Specimen Collection Kit. You may also submit a swab specimen using the Gen-Probe® Aptima Unisex Specimen Collection Kit for Endocervical and Male Urethral Swab Specimens. Both the urine and swab kits may be used for male and female patients. The Pap may only be used for female patients. Swab and urine kit specimens do not need to be refrigerated. Photos of these kits are included as Attachments 4 and 5.

Herpes

Summit Pathology sends requests for Herpes testing to Access Genetics, who report molecular detection of Herpes Simplex Virus, both Type 1 and Type 2. The specimen sample for this test can be taken from either the ThinPrep® or SurePath™ Pap vial. Those tests can be added to a pap that was sent to Summit Pathology for one month after the specimen was received.

If you are not taking a liquid based pap specimen, or the patient is male, the BBL™ CultureSwab™ Collection and Transport System may be used to obtain a swab specimen. A photo of this kit is included as Attachment 6. For best results, unroof the lesion before taking a specimen. This type of specimen does not need to be refrigerated.

Cystic Fibrosis

Genetic testing to identify Cystic Fibrosis carriers can be performed from a ThinPrep® or SurePath™ Pap specimen. This test is performed at a reference laboratory.

Strep B

Testing to identify Strep B can be completed from a liquid pap specimen.

Vaginosis Panel (Gardnerella, Trichomonas, Candida) and Tier 2 Pathogens Testing

Summit Pathology uses the only FDA approved method to test for Gardnerella, Trichomonas and Candida, the BD Affirm™ VPIII Microbial Identification Test. Specimens utilizing this method of testing must be submitted in the Affirm™ VPIII Ambient Temperature Transport System. Instructions for the collection of these specimens can be found in Attachment 7. A photo of the Affirm VPIII Ambient Temperature Transport System is included in Attachment 8. This specimen must be tested within 72 hours of specimen collection.

If no BD Affirm™ VPIII kit is sent, a pathogens panel testing for Gardnerella, Trichomonas and Candida can be performed from the pap vial. This testing is done at a reference laboratory. If you so choose, Tier 2 testing can be added, pending the outcome of the pathogens test. Tier 2 testing is ONLY from the pap vial. In Tier 2 testing, if the specimen is positive for Candida, a Candida Species profile is done. And if the specimen is negative for Gardnerella, a test for Atopobium vaginae is performed.

Non-Gynecologic Cytology

Specimens submitted to Summit Pathology should be labeled with two separate and unique identifiers, as required by the College of American Pathologists (CAP). All specimens must be accompanied by a Summit Pathology requisition form. If you are out of forms, please go to our website, www.summitpathology.com and go to the "Forms" tab. Print the requisition labeled "Cytology/Molecular/AP Requisition". Non-GYN cytology information is located on the bottom portion of the form. Requisition forms must be completed with patient name, date of birth, complete address and phone number, insurance information, date of specimen collection, physician name, appropriate clinical history, ICD9 code, site of specimen, and testing requested. Specimen containers should be placed into the main part of the plastic biohazard bags supplied by Summit Pathology. The requisition form should be placed into the outer pouch of the bag.

Body Fluids

Body fluid, such as ascites, peritoneal, pericardial and joint fluid should be submitted fresh, when obtained at a facility with pathology services on site, and should be refrigerated until pickup by a Summit authorized courier. In other cases, the fluid should be submitted in CytoLyt.

Breast/ Nipple Discharge

Should be submitted on an alcohol fixed or air dried slide

Brushes

Brushes, such as bronchial and ureteral should be fixed in CytoLyt to cover the specimen.

Bronchial Washings

Bronchial washings, including BAL should be submitted in CytoLyt.

Cerebrospinal Fluid

Submission of CSF specimens may include an air-dried Wright stained slide in addition to the fluid specimen sample. If the specimen is being submitted from 7 am Monday until 4 pm Friday, refrigerate the fresh specimen until pickup by a Summit authorized courier. If the specimen is being submitted after 4pm on Friday, submit the specimen in a vial of PreservCyt.

Fine Needle Aspiration

Fine needle aspirations from the breast, thyroid or other area should be submitted in CytoLyt Solution.

Sputum

Sputum should be submitted in CytoLyt Solution.

Tzank Smear (skin)

Tzank smears should be sumitted in 95% reagent alcohol or spray fixative

Urine

Urine should be submitted in CytoLyt Solution or 50% alcohol. Indicate which method was used to obtain the specimen (voided, catheter, bladder wash).

Biopsies

Specimens submitted to Summit Pathology should be labeled with two separate and unique identifiers, as required by the College of American Pathologists (CAP). All specimens must be accompanied by a Summit Pathology requisition form. If you are out of forms, please go to our website, www.summitpathology.com and go to the "Forms" tab. Print the requisition labeled "Surgical Pathology Requisition". Requisition forms must be completed with patient name, date of birth, complete address and phone number, insurance information, date of specimen collection, physician name, appropriate clinical history, ICD9 code, site of specimen, and testing requested. Specimen containers should be placed into the main part of the plastic biohazard bags supplied by Summit Pathology. The requisition form should be placed into the outer pouch of the bag.

Specimens for routine pathological evaluation should be submitted in 10% neutral buffered formalin. The amount of formalin should ideally be at least 10 times the volume of the specimen. A formalin health hazard label must be on each container (OSHA requirement). Be certain that small specimens do not get caught in the lid or the side of the container. Any breast tissue should have the in-formalin time noted on the requisition. All biopsies should be submitted in formalin with the exception of:

- Fresh or frozen section specimens
- Flow cytometry specimens
- Lymph node biopsies
- Muscle and nerve biopsies
- Renal biopsies
- Skin biopsies for immunofluorescence or electron microscopy
- Bone marrow biopsies for flow cytometry
- Specimens submitted for chromosome analysis
- Products of conception if cytogenetic analysis is requested

Fresh or Frozen Specimen Procedures

Specimens that are submitted fresh or for frozen section should only be taken from hospital or surgery center locations where a pathologist is available on call. Please follow the following procedure at the appropriate facility:

DURING BUSINEES HOURS (M-F 7:30AM – 5PM)

- North Colorado Medical Center
    - **FIRST** call 970-350-6400 (NCMC Lab Customer Service)

- If no immediate answer, call Summit Pathology 970-212-0530
- If no immediate answer, page the pathologist on call 800-920-6227

- McKee Medical Center
  - **FIRST** call 970-635-4163 (McKee Pathology Customer Service)
  - If no immediate answer, call 970-635-4133 (McKee Main Laboratory)
  - If no immediate answer, call Summit Pathology 970-212-0530
  - If no immediate answer, page the pathologist on call 800-920-6227

- Medical Center of the Rockies
  - **FIRST** call 970-624-2088 (MCR Gross Room)
  - If no immediate answer, page the pathologist on call 800-920-6227

- Poudre Valley Hospital
  - **FIRST** call 970-495-8740, option 1 (PVH pathology)
  - If no immediate answer, page the pathologist on call 800-920-6227

- Estes Park Medical Center
  - Call Summit Pathology at 970-212-0530 during business hours **ONLY** to schedule

- East Morgan County Hospital
  - Call Summit Pathology at 970-212-0530 during business hours **ONLY** to schedule

- Sterling Regional Medical Center
  - Call 970-521-3156 (SRMC pathology office) during business hours **ONLY** to schedule

- Colorado Plains Medical Center
  - Call Summit Pathology at 970-212-0530 during business hours **ONLY** to schedule

AFTER BUSINEES HOURS

- North Colorado Medical Center
  - **FIRST** call 970-350-6400 (NCMC Lab Customer Service)
  - If no immediate answer, page the pathologist on call 800-920-6227

- McKee Medical Center
  - **FIRST** call 970-635-4133 (McKee Main Laboratory)
  - If no immediate answer, page the pathologist on call 800-920-6227

- Medical Center of the Rockies
  - Page the pathologist on call 800-920-6227

- Poudre Valley Hospital
    - Page the pathologist on call 800-920-6227

Flow Cytometry Specimens

Flow cytometric studies require viable cells, therefore specimens should be submitted fresh, or in an appropriate fixative such as RPMI, not formalin. Please be a specific as possible in identifying the specific site of the biopsy, for example: "lymph node, left axilla". Appropriate clinical history should include CBC/ SPEP results, peripheral smear reviews, assessment for lymphadenopathy/ splenomegaly, 'B' symptoms, prior cytogenic studies, travel history, etc. The most common types of specimens submitted for flow include:

- Bone marrow aspirate, peripheral blood, lymph node biopsy material and body fluids such as pleural or cerebrospinal fluid.
- If lymphoma/ hematopoietic malignancy is a clinical possibility, the specimen should be submitted fresh.
- Bone marrow aspirate material and peripheral blood may be placed in green or purple top tubes. Please submit at room temperature, do not refrigerate. Please notify the lab if additional studies are requested (e.g. cytogenetics, specific markers to assess for myeloproliferative disease, FISH)
- Solid tissue, such as lymph nodes and body fluids may be submitted in sterile containers.

Lymph Node Biopsies

If lymphoma/ hematopoietic malignancy is a clinical possibility, submit the tissue fresh (without formalin) for possible flow cytometric studies. Lymph nodes submitted for disease process other than lymphoma/ leukemia (e.g. metastatic carcinoma) should be placed in a formalin container. Please be as specific as possible when identifying the specimen collection site and include appropriate clinical history, including CBC/ SPEP results, peripheral smear reviews, assessment for lymphadenopathy/ splenomegaly, 'B' symptoms, prior cytogenic studies, travel history, etc.

Muscle and Nerve Biopsies

Skeletomuscular pathology and peripheral nerve studies (for weakness) require special studies and expertise in these areas. Summit Pathology refers these cases to the University of Colorado for processing and assessment. Muscle biopsies should be 15 mm long with a diameter of 5-10 mm. Sutures around the width of the specimen help to orient it correctly.

Two (2) 20 mm long specimens should be submitted for nerve biopsies.

Both muscle and nerve biopsies should be wrapped in moist saline gauze. Do not float the specimen in saline.

Specimens should be delivered the day of the procedure, so advanced notice of a procedure is critical.

Renal Biopsies

Renal pathology is a highly specialized area, requiring several special techniques for full evaluation. Summit Pathology does not process renal biopsies for medical disease evaluation in our laboratory. We do process and evaluate biopsies for renal masses and tumors, however. Currently, Summit Pathology refers all medical renal biopsies to a reference laboratory for evaluation. Here are several general guidelines for these specimens:

- Proper evaluation of renal biopsies for medical disease requires evaluation of tissue under light microscopy, electron microscopy, and immunofluorescent histology. Each of these requires proper fixation of the biopsy material.
- Usually, the tissue is divided at the time of the procedure for proper preservation. The portion for light microscopy is placed in 10% formalin. The portion for electron microscopy is placed in 1% glutaraldehyde (Trumps fixative). The portion for immunofluorescent histology is placed in Zeus tissue fixative. All of these fluids are available as part of the Mayo Clinic Renal Biopsy kit, stocked by all our affiliated hospital laboratories.
- You will also need to complete the Renal Biopsy Patient Information Sheet. As these biopsies are sent to Mayo by our affiliated laboratories, please contact the appropriate hospital laboratory to arrange transport.

Skin Biopsies for Immunofluorescece

Currently, Summit Pathology forwards skin specimens to a reference laboratory for immunofluorescence studies. Specimens should be submitted in Zeus (also called Michael's) fixative or saline. These green top tubes are available from Summit Pathology. There is also a separate requisition form to be filled out for immunofluorescence studies. A separate specimen should also be submitted, in the same bag, for histologic studies done at Summit Pathology. Both specimen containers can be submitted in the same bag (with both requisition forms) to Summit Pathology.

Bone Marrow Biopsies

Please notify your hospital laboratory when a specimen is ready for submission. General guidelines for bone marrow biopsies are:

- Bone marrow cores are placed in formalin.
- A portion of the bone marrow aspirate should be placed in a green top tube(s) for possible flow cytometry and cytogenetic studies. Two tubes may be necessary if multiple studies are requested. Any remaining aspirate should be placed in formalin for 'clot sections'. Bone marrow particles for the aspirate smears are obtained from the green top tube.

- Unstained smears are made for cytochemical studies, when indicated (acute leukemia).
- **A CBC and peripheral smear should be ordered and performed within 24 hours of the procedure.**
- Please be specific with site of specimen. For example, if you want the report to state that the specimen was a posterior iliac crest biopsy rather vertebral biopsy, please let us know. Include appropriate clinical history, such as CBC results, electrophoresis, etc.
- **Please notify the hospital laboratory when the specimen is ready, as these biopsies require urgent handling.**

Chromosome Analysis

Chromosome analysis is most commonly performed on lymphoid material or bone marrow aspirate material/ peripheral blood. Lymphoid material should be submitted in RPMI. Peripheral blood and/or bone marrow aspirate should be submitted in a sodium heparin (green top) tube. These studies are performed at a reference laboratory.

Products of Conception

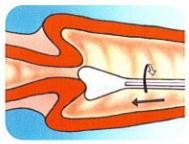
The procedure and handling of products of conception varies between hospital systems that are served by Summit Pathology. Please call your local clinical laboratory to determine the proper procedure.

Attachment 1

ThinPrep® Specimen Collection Guide

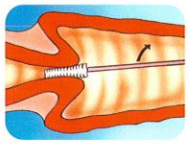
Quick Reference Guide
Endocervical Brush/Spatula Protocol

THE ThinPrep® PAP TEST



Obtain...
an adequate sampling from the ectocervix using a plastic spatula. If desired, use lukewarm water to warm and lubricate the speculum. Water-soluble gel lubricant sparingly applied to the posterior blade of the speculum can be used if necessary.[1] Select contoured end of plastic spatula and rotate it 360 degrees around the entire exocervix while maintaining tight contact with exocervical surface.

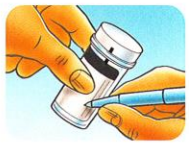
Rinse...
the spatula as quickly as possible into the PreservCyt® Solution vial by swirling the spatula vigorously in the vial 10 times. Discard the spatula.



Obtain...
an adequate sampling from the endocervix using an endocervical brush device. Insert the brush into the cervix until only the bottom-most fibers are exposed. Slowly rotate 1/4 or 1/2 turn in one direction. DO NOT OVER-ROTATE.

Rinse...
the brush as quickly as possible in the PreservCyt Solution by rotating the device in the solution 10 times while pushing against the PreservCyt vial wall. Swirl the brush vigorously to further release material. Discard the brush.

Tighten...
the cap so that the torque line on the cap passes the torque line on the vial.



Record...
the patient's name and ID number on the vial.

Record...
the patient information and medical history on the cytology requisition form.

Place...
the vial and requisition in a specimen bag for transport to the laboratory.

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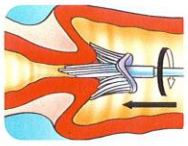
SIDE 1 OF 2

CERVICAL CANCER SCREENING

Quick Reference Guide
Broom-Like Device Protocol

THE ThinPrep® PAP TEST



Obtain...
an adequate sampling from the cervix using a broom-like device. If desired, use lukewarm water to warm and lubricate the speculum. Water-soluble gel lubricant sparingly applied to the posterior blade of the speculum can be used if necessary.[1] Insert the central bristles of the broom into the endocervical canal deep enough to allow the shorter bristles to fully contact the ectocervix. Push gently, and rotate the broom in a clockwise direction five times.

Rinse...
the broom as quickly as possible into the PreservCyt® Solution vial by pushing the broom into the bottom of the vial 10 times, forcing the bristles apart. As a final step, swirl the broom vigorously to further release material. Discard the collection device.

Tighten...
the cap so that the torque line on the cap passes the torque line on the vial.

Record...
the patient's name and ID number on the vial.

Record...
the patient information and medical history on the cytology requisition form.

Place...
the vial and requisition in a specimen bag for transport to the laboratory.

www.thinprep.com

1. Cervicovaginal Cytology Based on the Papanicolaou Technique; Approved Guideline – Third Edition (Clinical and Laboratory Standards Institute GP15-A3).

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SIDE 2 OF 2

Attachment 2

Lubricant use with the ThinPrep® Pap test

January 30, 2012

Re: Lubricant use during Pap sample collection

Dear Colleague,

On occasion, Hologic personnel are asked to provide information concerning the use of lubricants when collecting a Pap sample using the ThinPrep® Pap Test. As part of Hologic's continuing education for clinicians and laboratorians, this bulletin addresses the proper preparation of the cervix for an adequate Pap sample collection pertaining to the ThinPrep Pap Test and the use of lubricants on the speculum. Steps taken by the clinician, from patient education to improved sampling technique, may ensure that the sample collected maximizes the potential of the Pap test.[1,2]

Patient Education:

Women should be counseled to refrain from intercourse, douching, using tampons, or using intravaginal medication for at least 48 hours before the examination to decrease the possibility that the number of exfoliated cells will be diminished or obscured by personal lubricants or spermicides.[1,2] In addition, the patient should avoid scheduling her appointment during heavy menstrual bleeding.[1] If you would like Hologic patient education materials for your office, please visit www.hologiccustomersolutions.com.

Sample Collection Options for Lubricating the Speculum:

1. **Lukewarm Water:** For a patient without physical or physiologic reasons for needing lubricant, lukewarm water may be used to warm and lubricate the speculum. This protocol has the least risk to the quality of the Pap sample collected. [1,3] Professional organizations including ACOG and CLSI recognize that excessive use of lubricant may contaminate or obscure the Pap sample.

2. **Lubricant Gels:** If lubricant must be used due to patient discomfort or other circumstances, lubricant should be used sparingly and applied ***only*** to the exterior sides of the speculum blades, ***avoiding contact with the tip of the speculum***.[1,2,3,4] (see pictures below) When a lubricant is used sparingly and appropriately, it poses little risk to the quality of the Pap sample. However, when a lubricant is used in excess, it can adversely affect the Pap sample. Hologic evaluated a variety of popular lubricants and found those containing carbomer or carbopol polymers (thickening agents) interfere with the ThinPrep Pap test when found in the sample vial.[5] Hologic recognizes the varying availability of different types of lubricants and recommends that, if used, any lubricant should be applied sparingly as described below.

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Appropriate Use of Lubricant for Pap Collection

Lubricant gels should be used sparingly.

Apply only to exterior sides of the speculum, avoiding the tip.

Should you have further questions regarding this topic, please refer to the CLSI guidelines or contact Hologic Technical Support Department at 1-800-442-9892, option 6.

Sincerely,

Edward Evantash
Medical Director

1. Davey *et al*, 2008, "Cervical Cytology Specimen Adequacy: Patient Management Guidelines and Optimizing Specimen Collection"; American Society for Colposcopy and Cervical Pathology Journal of Lower Genital Tract Disease, Volume 12, Number 2, 2008, 71-81
2. Amies, AE.; Miller, L; Lee, Shu-Kuang; Koutsky, L, The Effect of Vaginal Speculum Lubrication on the Rate of Unsatisfactory Cervical Cytology Diagnosis, Obstet Gynecol. 100(5, Part 1):889-892, November 2002.
3. "Cervicovaginal Cytology Based on the Papanicolaou Technique; Approved Guideline – Third Edition", Clinical and Laboratory Standards Institute (formerly NCCLS), Vol. 28 No. 28, 2008.
4. ACOG Practice Bulletin, Clinical Management Guidelines for Obstetrician Gynecologists, Number 109, December 2009, pg 2.
5. Hologic internal study, Data on file.

MISC-00579-001 Rev. 004

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www.hologic.com

Attachment 3

SurePath™ Specimen Collection Guide

Specimen Collection for the BD SurePath™ Pap test

1. Collect

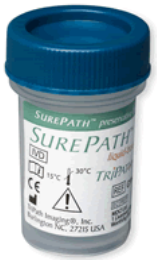
Collect the cytology sample using either a broom-like device, or combination brush/spatula with detachable heads. Follow manufacturer collection instructions for detachable head device(s).

2. Drop

Drop the detachable head device(s) into the BD SurePath™ vial.

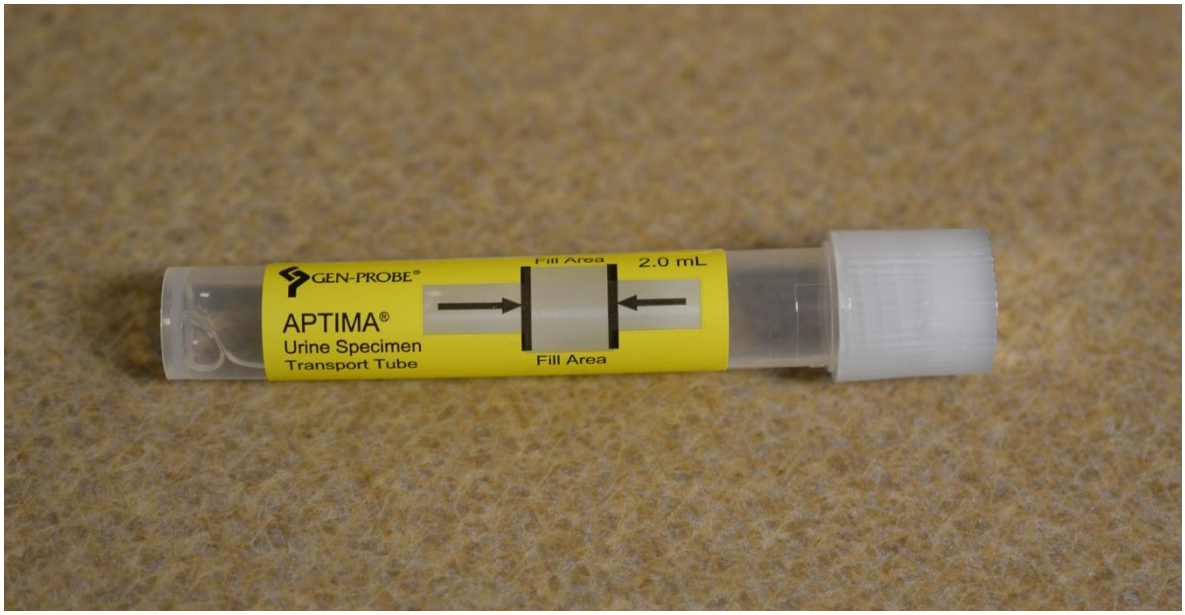
3. Send

Place the cap on the vial and tighten. Send the BD SurePath™ vial to the lab for processing.



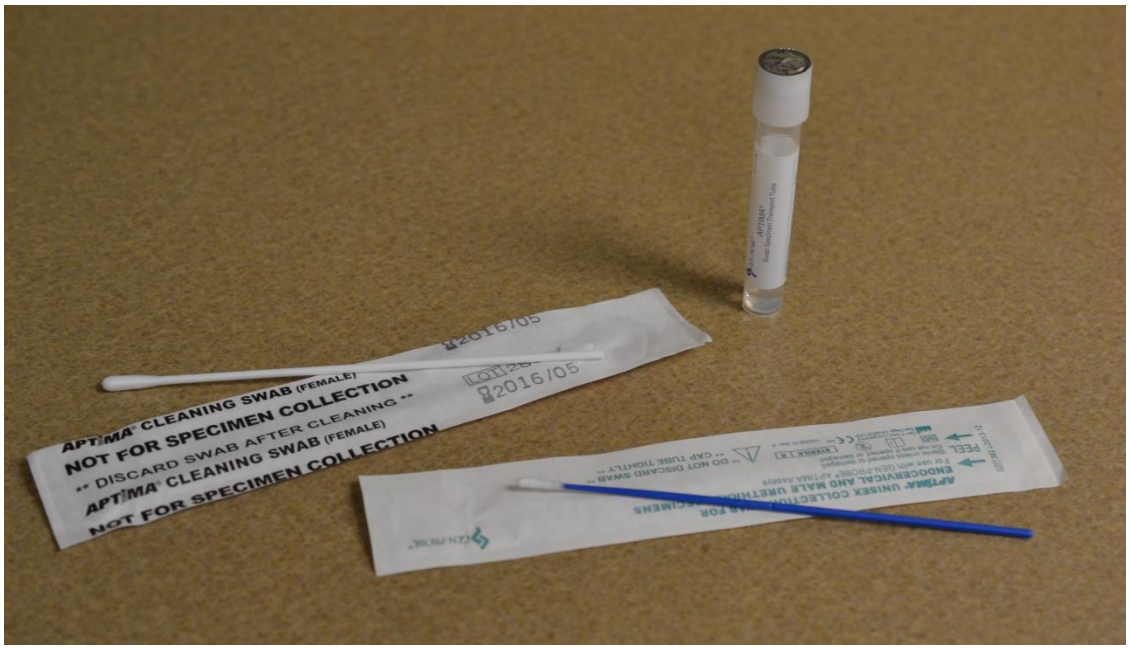
Attachment 4

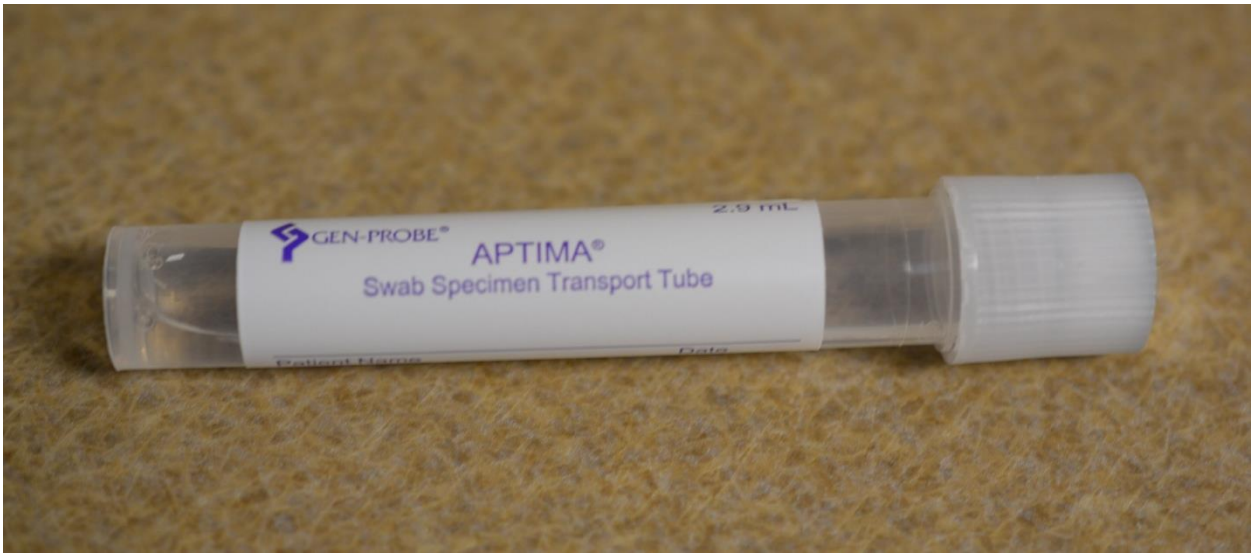
Gen-Probe® Aptima Urine Specimen Collection Kit


GEN-PROBE®
APTIMA®
Urine Specimen
Transport Tube
Fill Area
2.0 mL
Fill Area

Attachment 5

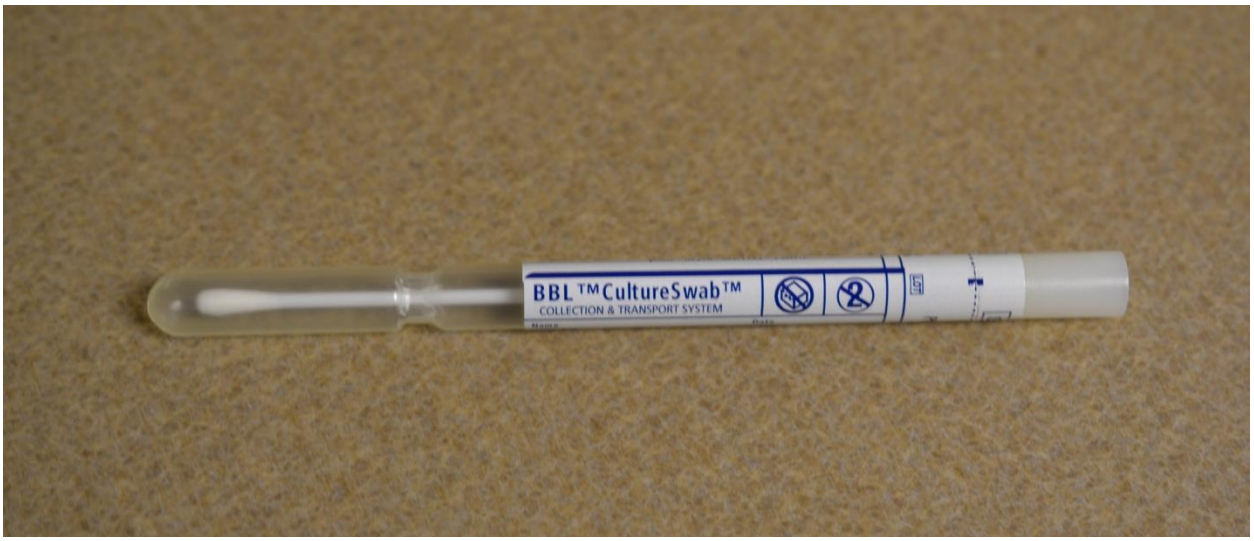
Gen-Probe® Aptima Unisex Swab Specimen Collection Kit


APTIMA CLEANING SWAB (FEMALE)
NOT FOR SPECIMEN COLLECTION
** DISCARD SWAB AFTER CLEANING **
APTIMA CLEANING SWAB (FEMALE)
NOT FOR SPECIMEN COLLECTION
2016/05


GEN-PROBE
APTIMA
Swab Specimen Transport Tube
2.9 mL

Attachment 6

BBL™ CultureSwab™ Collection and Transport System


BBL™ CultureSwab™
COLLECTION & TRANSPORT SYSTEM

Attachment 7

BD Affirm™ VPIII Microbial Identification Test Collection Guide
(Vaginosis Panel: Gardnerella, Trichomonas, Candida)

BD Affirm™ VPIII Collection and Transport

BD Affirm™ VPIII Ambient Temperature Collection System

For collection of vaginal specimens for use in the BD Affirm™ VPIII Microbial Identification Test for *Candida, Gardnerella* and *Trichomonas*. See reverse side for transport procedures.

VAGINAL SAMPLE COLLECTION

Sample collection is a critical step. Personnel collecting vaginal fluid specimens should be well trained to ensure adequate sample collection. **All samples must be collected using the materials (swabs and tubes) provided in the set.**

1. Place the patient in position for a pelvic examination. Insert a speculum into the vagina to permit visualization of the posterior vaginal fornix.
2. Using the sterile swab, obtain a sample from the posterior vaginal fornix. Twist or roll the swab against the vaginal wall two or three times, ensuring the entire circumference of the swab has touched the vaginal wall. Swab the lateral vaginal wall while removing the swab. (see Figure 1)
3. Immediately place the swab in the Sample Collection Tube (SCT).
4. With the swab touching the **BOTTOM** of the collection tube, grasp the pre-scored handle of the swab just above the top of the tube and bend until the swab breaks (Figure 2). When the swab is fully inserted into the collection tube, the score mark on the swab is approximately 1 cm above the top of the collection tube. Discard the broken handle into an infectious waste container.
5. Place the cap over the exposed end of the swab and firmly press the cap onto the tube. The cap will "snap" onto the tube when it is properly seated.
6. Label the **Sample Collection Tube (SCT)** with the patient identification information. Include the time the sample was collected.
7. Place the capped **Sample Collection Tube (SCT)** into the plastic **Sample Transport Bag** for transport for testing with the **Affirm** VPIII Microbial Identification Test.

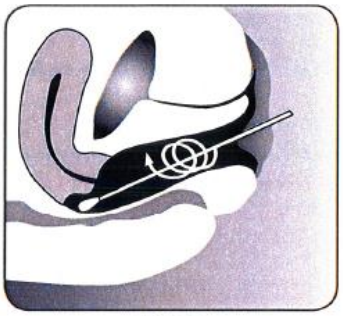


Figure 1

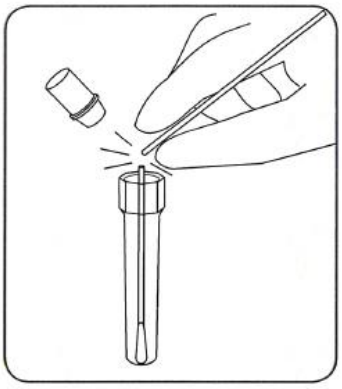


Figure 2

BD

Helping all people live healthy lives

BD Affirm™ VPIII Collection and Transport

BD Affirm™ VPIII Ambient Temperature Transport System

For transport of vaginal specimens for use in the BD Affirm™ VPIII Microbial Identification Test for *Candida*, *Gardnerella* and *Trichomonas*. See reverse side for collection procedures.

TRANSPORT

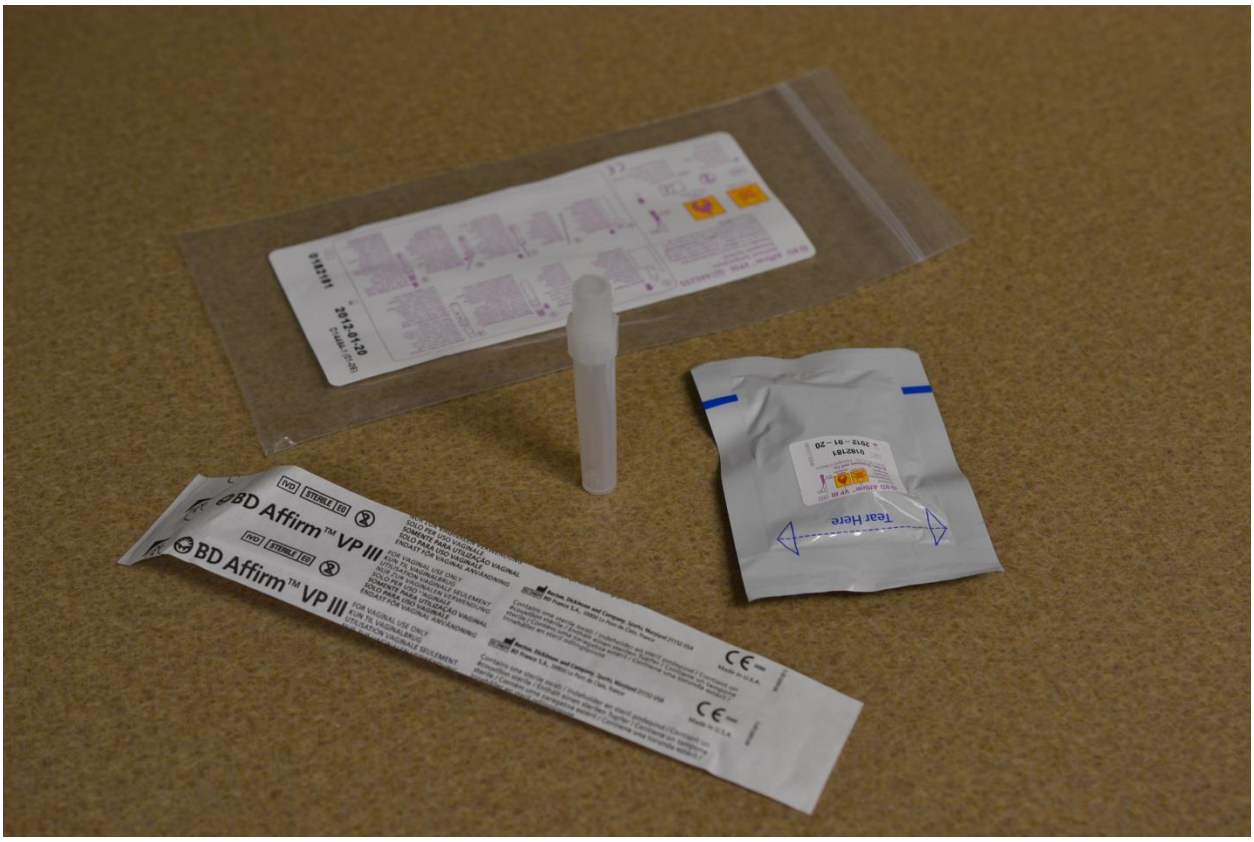
Sample is stable for 72 hours at ambient temperature (15-30°C) or refrigerated (2-8°C) storage.

BD

BD Diagnostics
7 Loveton Circle
Sparks, MD 21152-0999
800.638.8663
www.bd.com/ds

Attachment 8

BD Affirm™ VPIII Ambient Temperature Transport System


BD Affirm™ VP III
BD Affirm™ VP III
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